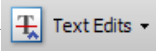
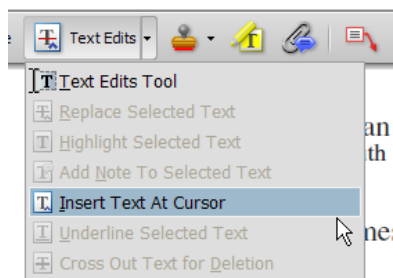


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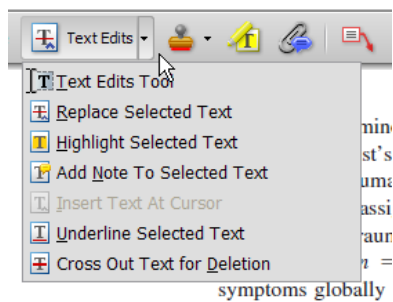
To facilitate electronic transmittal of corrections, we encourage authors to utilize the comments and notes features in Adobe Acrobat. The PDF provided has been “comment-enabled,” which allows you to utilize the comments and notes features, even if using only the free Adobe Acrobat reader (see note below regarding acceptable versions). Adobe Acrobat’s Help menu provides additional details on the tool. When you open your PDF, the comments/notes/edit tools are clearly shown on the tool bar (though icons may differ slightly among versions from what is shown below).

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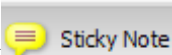
- Use the **Text Edits tool** () to insert, replace, or delete text.
 - To **insert text**, place your cursor at a point in text and select “Insert Text at Cursor” from the text edits menu. Type your additional text in the pop-up box.



- To **replace text** (do this instead of deleting and then re-inserting), highlight the text to be changed, select “Replace Selected Text” from the text edits menu, and type the new text in the pop-up box.



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- Use the **Sticky Note tool** () to describe changes that need to be made (e.g., changes in bold, italics, or capitalization use; altering or replacing a figure) or to answer a question or approve a change that was posed by the editor. To use this feature, click on the sticky note tool and then click on a point in the PDF where you would like to make a comment, then type your comment in the pop-up box.



- Use the **Callout tool** () to point directly to changes that need to be made. Try to put the callout box in an area of white space so that you do not obscure the text, as in the example below.

Table 5
Experiment 4: Comparative Optimism as a Function of Self-Presentation and Event Valence

	Event					
	Positive		Negative		Total	
Self-presentation	<i>M</i>	<i>SD</i>	<i>M</i>	<i>SD</i>	<i>M</i>	<i>SD</i>
Public/student	3.46	0.13	3.60	0.10	3.53	0.12
Public/expert	2.66	0.12	2.78	0.13	2.73	0.13
Control	2.39	0.11	2.46	0.09	2.43	0.11
Total	2.84	0.47	2.95	0.50		

The first column's entries should be flush left (except for "Total", which should be indented one em-space), as in Tables 1 and 2 previously.

- Use the **Highlight tool** () to indicate font problems, bad breaks, and other textual inconsistencies. Describe the inconsistencies with the callout tool (shown) or a sticky note. One callout (or sticky note) can describe many changes.

$$du/dt = -\lambda v^\alpha = -\lambda u$$

$$du/u = -\lambda dt$$

$$u_t = ue^{-\lambda t}$$

Close up minus sign to lambda (3 times, highlighted)

An alternate method is to select the appropriate text with your cursor, select “**Add Note to Selected Text**” from the text edits menu, and then type your note in the pop-up box (the selected text is highlighted automatically).

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Complete Genome Sequence of *Capripoxvirus* Strain KSGP 0240 from a Commercial Live Attenuated Vaccine

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Capripoxviruses cause economically important diseases in domestic ruminants in regions endemic for these viruses. We report here the complete genome sequence of the KSGP 0240 vaccine strain from the live attenuated vaccine Kenyavac (JOVAC).

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Viruses belonging to the *Capripoxvirus* genus, namely, goatpox virus (GTPV), sheeppox virus (SPPV), and lumpy skin disease virus (LSDV), cause important diseases in goats, sheep, and cattle, respectively. Vaccination plays a crucial role in limiting the diseases in regions that are endemic for the viruses. In these regions, the Kenyan SPPV and GTPV KSGP 0240 strain are widely used for vaccination against GTPV and SPPV. Although the virus was isolated from sheep, phylogenetic analyses of partial sequences have identified KSGP 0240 as a lumpy skin disease virus (1, 2). Here, we determined the complete sequence of the KSGP 0240 strain from the Kenyavac vaccine (JOVAC) directly from a commercial vaccine batch.

DNA was purified from a freeze-dried vaccine pellet dissolved in 3 ml of phosphate-buffered saline (PBS) using the Puregene core kit A (Qiagen), according to the manufacturer's instructions. Presequencing enrichment was performed through an in-house long-range PCR methodology covering the entire genome with overlapping ~5.5-kb amplicons. P6-C4 sequencing was performed on 1 single-molecule real-time (SMRT) cell on a PacBio RSII sequencer (Pacific Biosciences) at the Genomics Core UZ Leuven (Leuven, Belgium).

Consensus amplicon sequences were obtained from the reads using the LAA-protocol (default parameters; Pacific Biosciences) in SMRT Portal (Pacific Biosciences) version 2.3.0. These amplicons were further assembled into a unique contig using the iAssembler software (3). Discrepancies with previously published LSDV genomes were confirmed by Sanger sequencing. The protein-coding genes were predicted by NCBI ORF-Finder (<http://www.ncbi.nlm.nih.gov/orffinder/>) and by GATU relative to the Neethling NI-2490 reference strain (accession no. NC_003027) (4).

Consensus amplicon sequences assembled into a single double-stranded linear DNA sequence of 150,663 bp, with an average G+C content of 25.90%. The strain KSGP 0240 included in the Kenyavac vaccine contains a 145,938-bp central coding region flanked by two identical terminal inverted repeats of at least 2,192 bp. The KSGP 0240 genome shares 99.9% homology with

the LSDV strain Neethling NI-2490, differing by only a single amino acid substitution (P/Q in LSDV049) and a single-nucleotide deletion resulting in a frameshift that truncates the LSDV134 putative gene. Based on the complete genome sequence, the strain KSGP 0240 was confirmed to be an LSDV.

Accession number(s). The complete genome sequence of LSDV strain KSGP 0240 (Kenyavac vaccine) has been deposited in GenBank under accession number [KX683219](https://www.ncbi.nlm.nih.gov/nuccore/KX683219).

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