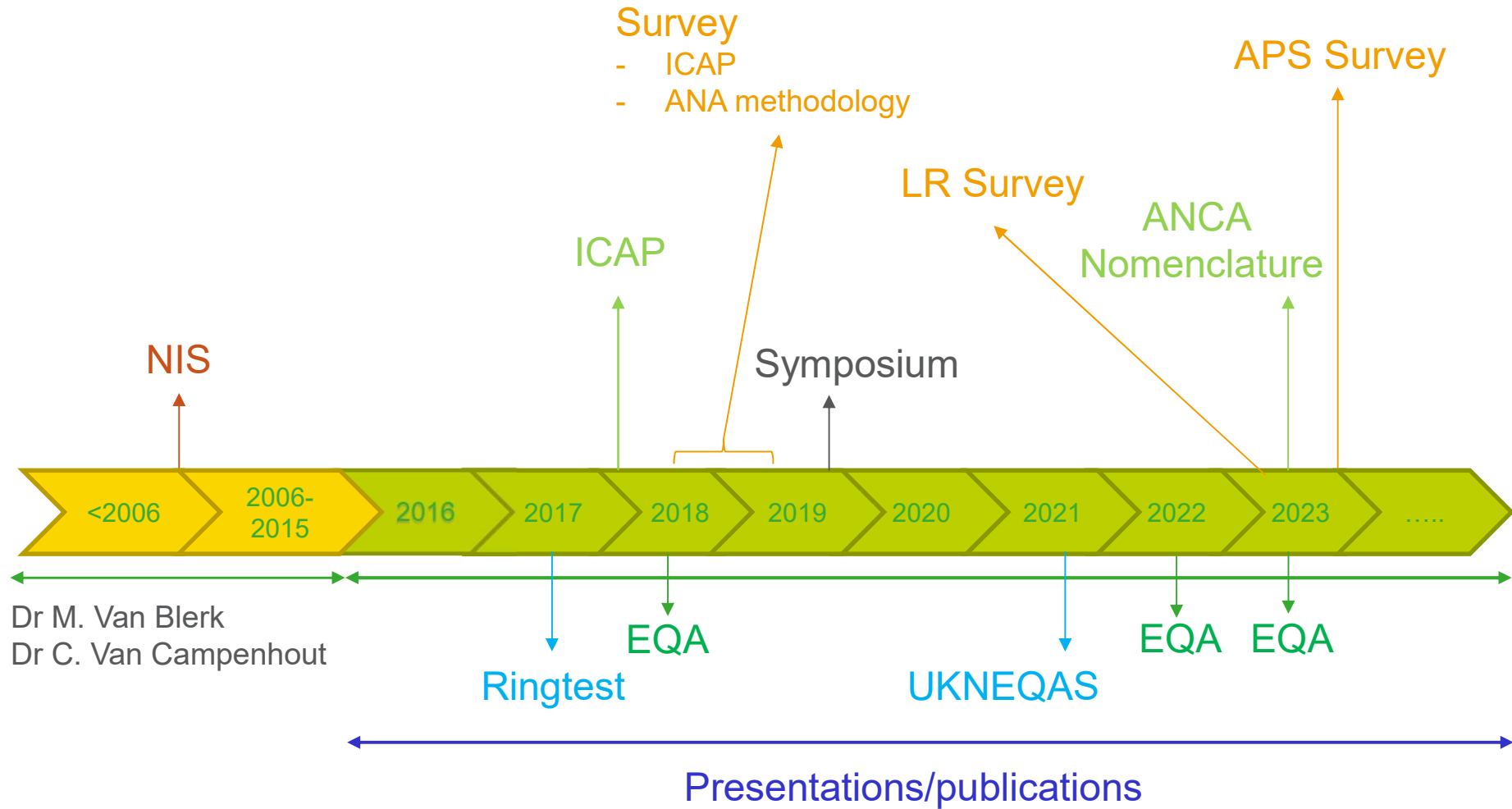


NIS EQA: AN OVERVIEW AND PERSPECTIVES

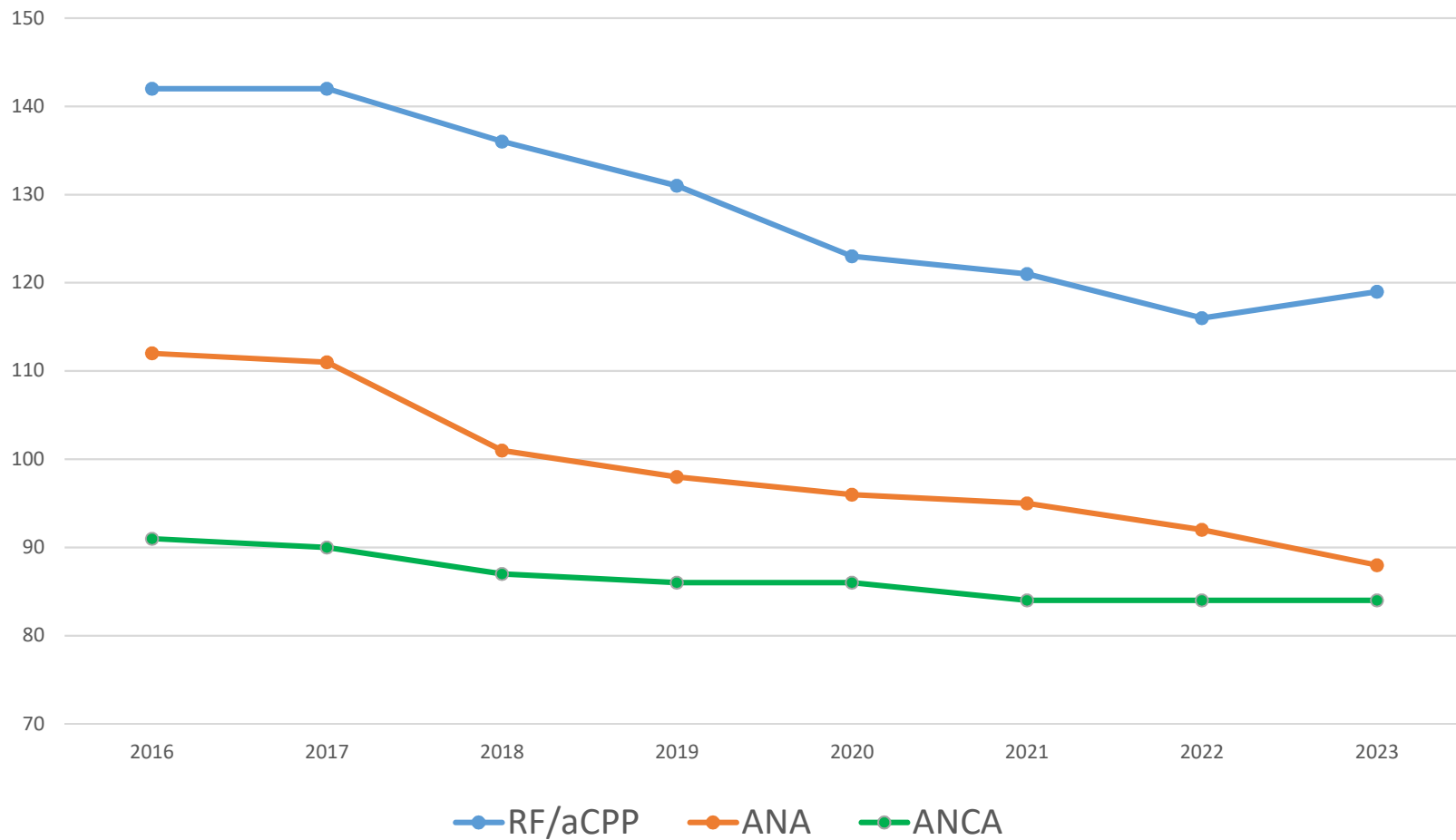
Broeders Sylvia
Sciensano

Symposium Non-infectious serology 08/03/2024

Timeline



EQA: Participants



EQA: RF/anti-CCP

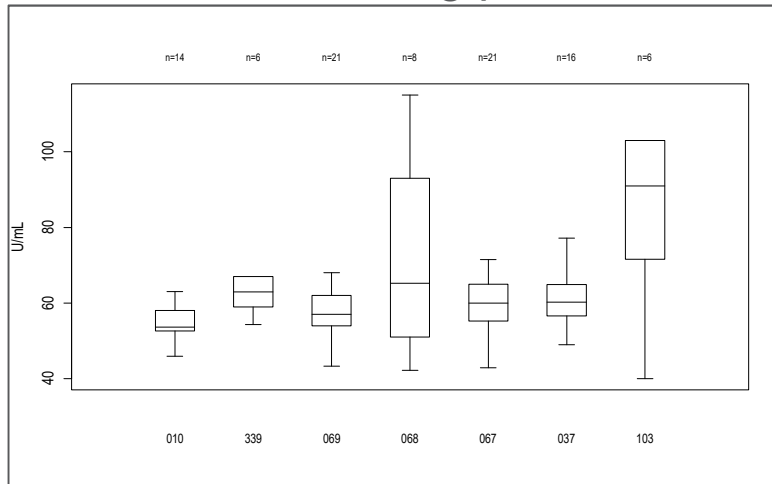
EQA	Sample nr	RF	Anti-CCP
2016-2	SN/14322	+	-
2017-1	SN/14479	Strong +	Strong +
2018-1	SN/15559	Weak +	Strong +
2019-1	SN/15969	Strong +	Strong +
2020-1	SN/16984	+	Strong +
2021-1	SN/17805	-	Strong +
2022-1	SN/18837	+	Strong +
2023-1	SN/19347	+	-

↓
1 sample for 2 EQA: RF/anti-CCP + ANA

RF/anti-CCP: Take-home messages

Method variation

RF: usually overlap/concordance between methods
 BUT weak >< strong pos



Results > range: dilute or not?

- Do like in routine
- Follow the insert of the kit

SN/19347

Anti-CCP: higher variation – Roche!

	2021	2022	2023
Abbott	107.8	143	0.6
Roche	496	>500	38
Siemens	72	74	<1.5
Inova		178	<1.2
ThermoSc	155	373	2.0
	St+	St+	-

Cut-off 17
 => +

!! Interference of anti-streptavidin antibodies: known but rare (<0.7%)

ACR/EULAR 2010 criteria !!

2010 Rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative

Daniel Aletaha,¹ Tuhina Neogi,² Alan J Silman,³ Julia Funovits,¹ David T Felson,² Clifton O Bingham III,⁴ Neal S Birnbaum,⁵ Gerd R Burmester,⁶ Vivian P Bykerk,⁷ Marc D Cohen,⁸ Bernard Combe,⁹ Karen H Costenbader,¹⁰ Maxime Dougados,¹¹ Paul Emery,¹² Gianfranco Ferraccioli,¹³ Johanna MW Hazes,¹⁴ Kathryn Hobbie,¹⁵ Tom WJ Huizinga,¹⁶ Arthur Kavanaugh,¹⁷ Jonathan Kay,¹⁸ Tore K Kvien,¹⁹ Timothy Laing,²⁰ Philip Mease,²¹ Henri A Ménard,²² Larry W Moreland,²³ Raymond L Naden,²⁴ Theodore Pincus,²⁵ Josef S Smolen,¹ Ewa Stanislawski,²⁶ Deborah Symmons,²⁷ Paul P Tak,²⁸ Katherine S Upchurch,¹⁸ Jiří Vencovsky,² Frederick Wolfe,³⁰ Gillian Hawker,³¹

Menarini Diagnostics

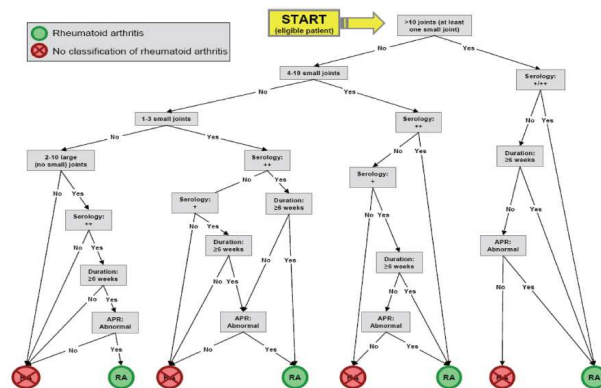
- Zenit RA CCP (CLIA) -> AU/ml
- Zenit CCP ELISA -> U/ml, ratio

- Use a quantitative method
 - Correct toolkit + units!
- Analyse RF and anti-CCP
- Check cut-off

Table 3 The 2010 American College of Rheumatology/European League Against Rheumatism classification criteria for RA

	Score
Target population (Who should be tested?): Patients who	
1) have at least 1 joint with definite clinical synovitis (swelling)*	
2) with the synovitis not better explained by another disease†	
Classification criteria for RA (score-based algorithm: add score of categories A-D; a score of ≥6/10 is needed for classification of a patient as having definite RA)‡	
A. Joint involvement§	
1 large joint¶	0
2–10 large joints	1
1–3 small joints (with or without involvement of large joints)**	2
4–10 small joints (with or without involvement of large joints)	3
>10 joints (at least 1 small joint)††	5
B. Serology (at least 1 test result is needed for classification)‡‡	
Negative RF <i>and</i> negative ACPA	0
Low-positive RF <i>or</i> low-positive ACPA	2
High-positive RF <i>or</i> high-positive ACPA	3
C. Acute-phase reactants (at least 1 test result is needed for classification)§§	
Normal CRP <i>and</i> normal ESR ¶¶	0
Abnormal CRP <i>or</i> normal ESR ¶¶	1
D. Duration of symptoms¶¶¶	
<6 weeks	0
≥6 weeks	1

a score of 6 or greater being indicative of the presence of definite RA. **.be**



EQA: ANCA

EQA	Sample nr	IIF	IA
2016-1	SN/14072	pANCA	MPO
2017-3	SN/14739	pANCA	MPO
2018-2	SN/15785	cANCA	PR3
	SN/15624	-	PR3
2019-2	SN/16467	pANCA	MPO
2020-2	SN/14741	pANCA	MPO
2021-2	SN/18126	cANCA	PR3
2022-2	SN/19094	pANCA	MPO
2023-3	SN/19999	cANCA	PR3

Publication

Nomenclature

Didactic sample

ANCA: Take-home messages

Variability/Sensitivity IIF vs IA

Revised 2017 international consensus on testing of ANCAs in granulomatosis with polyangiitis and microscopic polyangiitis

Xavier Bossuyt¹, Jan-Willem Cohen Tervaert², Yoshihiro Arimura³, Daniel Blockmans⁴, Luis Felipe Flores-Suárez⁵, Loïc Guillevin⁶, Bernhard Hellmich⁷, David Jayne⁸, J. Charles Jennette⁹, Cees G. M. Kallenberg¹⁰, Sergey Moiseev¹¹, Pavel Novikov¹¹, Antonella Radice¹², Judith Anne Savige¹³, Renato Alberto Sinico¹⁴, Ulrich Specks¹⁵, Pieter van Paassen¹⁶, Ming-hui Zhao¹⁷, Niels Rasmussen¹⁸, Jan Damoiseaux¹⁹ and Elena Csernok⁷

⇒ high-quality immunoassays can be used as the primary screening method for patients suspected of having the ANCA-associated vasculitides GPA and MPA without the categorical need for IIF

EQA SN/14739

Revised 2017 international consensus on ANCA testing in small vessel vasculitis: support from an external quality assessment

Sylvia Broeders¹, Sylvie Goletti², Jean-Paul Tomasi², Carolien Bonroy³, René-Louis Humbel⁴, Laurence Lutteri⁵, Sofie Schouwers⁶,  Lieve Van Hoovels⁷, Martine Vercaammen⁸, Xavier Bossuyt⁹

SN/15624: didactic sample

- Patient GPA under treatment: IIF neg, PR3
- High variability seen in both IIF and IA
 - > Lower titers & epitope spreading

>< RIZIV/INAMI
nomenclature

ANCA: nomenclature 2023

SN/19999

- Follow-up sample
- GPA patient
- cANCA-PR3
- 59/84 IIF -> OK
- 81/84 IA -> OK

Questions

- Have you adjusted your testing strategy for ANCA vasculitides in response to the new nomenclature?
- If not, do you plan to adjust your test strategy?

⇒ 80/84 answers

Adapted	To do	#	%
Y	N	43	53.8
Y(part)	Y	2	2.5
N	Y	23	28.8
N	N	12	15.0

CHANGE
68/80!
85%

??

EQA: not possible to give the whole history of the follow-up sample -> MPO/PR3
routine: IIF + IA -> also in EQA: make correlation!

EQA: ANA

	EQA	Sample nr	IIF	Anti-dsDNA	Anti-ENA
	2016-3	SN/14565	centromere	-	CENP-A/B
Ringtest	2017-2	SN/15085	(fine) speckled	-	SSA/SSB
		SN/15086	-	-	-
	2018-3	SN/15843	AC-1	+	+
	2019-3	SN/16557	AC-4	-	SSA/SSB
	2020-3	SN/531	AC-3	-	CENP-A/B
UKNEQAS	2021-3	SN/18676	AC-5	-	RNP
		SN/18677	AC-5	/	/
		SN/18647	AC-2	/	/
	2022-3	SN/19448	AC-3	-	CENP-B
		SN/19350	AC-22	/	/
	2023-2	SN/19347	AC-4	-	SSA

ICAP

Didactic sample

1 sample for 2 EQA: RF/anti-CCP + ANA

ANA: Take-home messages

SN/19448

- Centromere AC-3 -> CENP-A/B
 - Additional borderline: SSB, Ro52, gp210, Th/To, AMA-M2
- > sensitivity/specificity of methods!

Understanding and interpreting antinuclear antibody tests in systemic rheumatic diseases

Xavier Bossuyt¹, Ellen De Langhe^{2,3}, Maria Orietta Borghi^{4,5} and Pier Luigi Meroni⁴

Abstract | Antinuclear antibodies (ANAs) are valuable laboratory markers to screen for and support the diagnosis of various rheumatic diseases (known as ANA-associated rheumatic diseases). The importance of ANA testing has been reinforced by the inclusion of ANA positivity as an entry criterion in the 2019 systemic lupus erythematosus classification criteria. In addition, specific ANAs (such as antibodies to Sm, double-stranded DNA (dsDNA), SSA/Ro60, U1RNP, topoisomerase I, centromere protein B (CENPB), RNA polymerase III and Jo1) are included in classification criteria for other rheumatic diseases. A number of techniques are available for detecting antibodies to a selection of clinically relevant antigens (such as indirect immunofluorescence and solid phase assays). In this Review, we discuss the advantages and limitations of these techniques, as well as the clinical relevance of the differences between the techniques, to provide guidance in understanding and interpreting ANA test results. Such understanding not only necessitates insight into the sensitivity and specificity of each assay, but also into the importance of the disease context and antibody level. We also highlight the value of titre-specific information (such as likelihood ratios).

- The performance characteristics of IIF assays and SPAs are disease-dependent; IIF assays are more sensitive than SPA for screening for systemic sclerosis (and systemic lupus erythematosus) but not Sjögren syndrome.

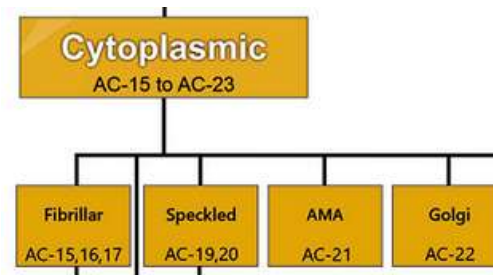
Choose well!!!

ANA: Take-home messages

SN/19350

- Didactic sample
- Golgi-like pattern AC-22
 - 93.3 % IIF +
 - 83.3 % correct pattern

- ICAP competent level



- No clinical association

Anti-Golgi autoantibodies are not clinically associated with systemic autoimmune diseases

Pieter Vermeersch,¹ Karolien Van den Bergh,¹ Daniel Blockmans,² Rene Westhovens,³ Xavier Bossuyt¹

ANA: Take-home messages

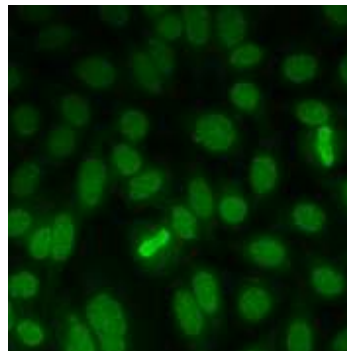
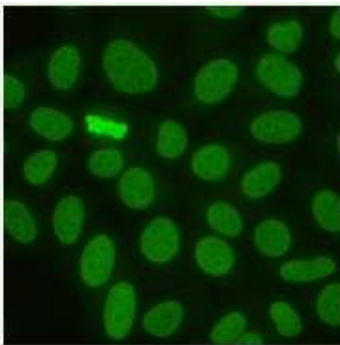
SN/19347 – SN/16557 – SN/18647

IIF:

- SSA on HEp-2000

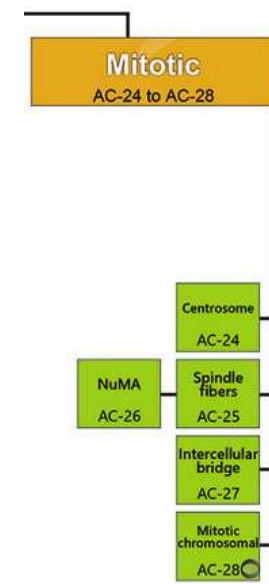
Note #1. Anti-SS-A/Ro60 staining on HEp-2000 slides (IMMUNO CONCEPTS) is different from other HEp-2 slides.

- Homogeneous (AC-1) and DFS70 (AC-2): Nuclear - positive mitosis



<https://www.anapatterns.org/trees-2021.php>

≠ mitotic patterns (AC-24->28)



ANA: Take-home messages

SN/19347

Anti-dsDNA:

Different methodologies (buffer, Ag source, conjugate,...)

- > difference performance
- > difference in detection of Ab

Belgian recommendations on ANA, anti-dsDNA and anti-ENA antibody testing

M. Van Blerk^{1,2}, X. Bossuyt^{2,3}, R. Humbel^{2,4}, A. Mewis^{2,5}, G. Servais^{2,6}, J. P. Tomasi^{2,7}, C. Van Campenhout^{1,2}, L. Van Hoovels^{2,8}, M. Vercammen^{2,9}, J. Damoiseaux¹⁰, W. Coucke¹, P. Van de Walle¹

- > choice of cut-off
 - 97.4 %: dsDNA neg
 - Diesse (cut-off: 20)

=> It is not recommended to use a dot technique for anti-dsDNA antibody detection.

=> The method used for determining anti-dsDNA antibodies should be specified when reporting the result.

Result	Cut-off	Conclusion
18.8 IU/ml	<10	+

ANA: Take-home messages

SN/19347

Anti-ENA:

Fine speckled AC-4 – SSA/Ro60

- 57/58 dot -> 1/58 Ro52 only tested
- 22/22 ELISA/CLIA/FEIA
- 4/4 microarray

-> Need to test basic Ag

-> Adapt method of IIF

-> Dot/line: specificity

=> Correlation IIF/anti-ENA

Belgian recommendations on ANA, anti-dsDNA and anti-ENA antibody testing

M. Van Blerk^{1,2}, X. Bossuyt^{2,3}, R. Humbel^{2,4}, A. Mewis^{2,5}, G. Servais^{2,6}, J. P. Tomasi^{2,7}, C. Van Campenhout^{1,2}, L. Van Hoovels^{2,8}, M. Vercammen^{2,9}, J. Damoiseaux¹⁰, W. Coucke¹, P. Van de Walle¹

It is recommended to type the anti-ENA antibodies for at least :

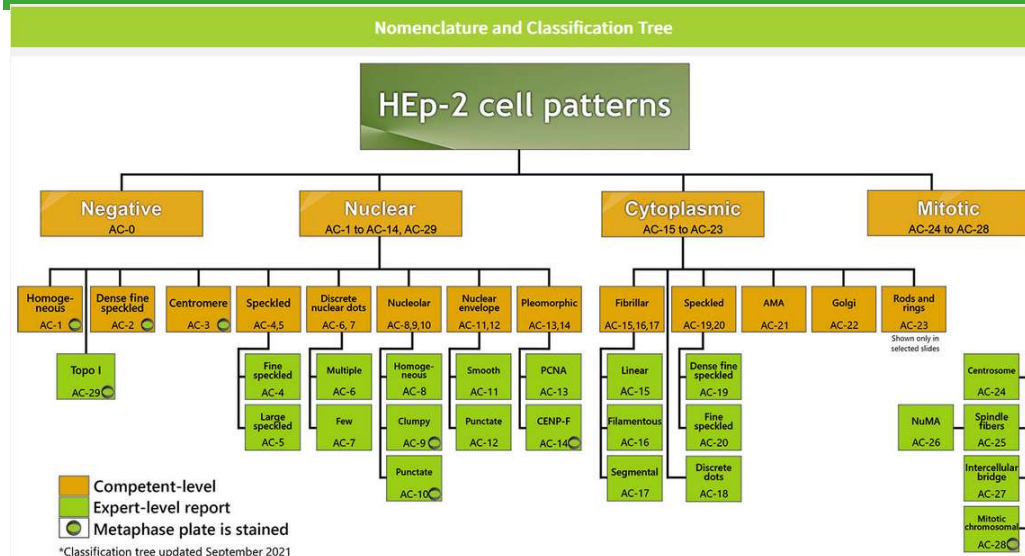
SSA/Ro60, SSB, Sm, RNP, CENP-B, Scl-70, Jo-1

Carolien Bonroy, Martine Vercammen, Walter Fierz, Luis E.C. Andrade, Lieve Van Hoovels, Maria Infantino, Marvin J. Fritzler, Dimitrios Bogdanos, Ana Kozmar, Benoit Nespola, Sylvia Broeders, Dina Patel, Manfred Herold, Bing Zheng, Eric Y.T. Chan, Raivo Uibo, Anna-Maija Haapala, Lucile Musset, Ulrich Sack, Gabor Nagy, Tatjana Sundic, Katarzyna Fischer, Maria-José Rego de Sousa, Maria Luisa Vargas, Catharina Eriksson, Ingmar Heijnen, Ignacio García-De La Torre, Orlando Gabriel Carballo, Minoru Satoh, Kyeong-Hee Kim, Edward K.L. Chan, Jan Damoiseaux, Marcos Lopez-Hoyos and Xavier Bossuyt* for the European Federation of Laboratory Medicine (EFLM) Working Group "Autoimmunity Testing," the European Autoimmune Standardization Initiative (EASI) and International Consensus on Antinuclear Antibody Patterns (ICAP)

**Detection of antinuclear antibodies:
recommendations from EFLM, EASI and ICAP**

09:30 – 09:55 Detection of antinuclear antibodies: recommendations from EFLM, EASI and ICAP: clinical and methodological aspects including automated microscopy
Carolien Bonroy (Department of Diagnostic Sciences, Ghent University, Ghent, Belgium;
Department of Laboratory Medicine, University Hospital Ghent, Ghent, Belgium)

ANA: ICAP nomenclature 2018 -> 2021



<https://www.anapatterns.org/trees-2021.php>

ICAP Smartphone App – connecting to our new generation of users

International Consensus on ANA Patterns (ICAP): inbedding in het Nederlandse taalgebied

Jan Damoiseaux, Xavier Bossuyt, Sylvia Broeders, Dörte Hamann, Karina de Leeuw, Maarten Limper, Henny Otten, Caroline Roozendaal, Sofie Schouwers, Marco Schreurs, Rogier Thurlings, Paul van Daele, Renate van der Molen, Lieve van Hoovels, Martine Vercammen

SN/15843: EQA

Vertical in the window

1	2	3	4
<input type="checkbox"/> NEGATIEF (AC-0)			
<input type="checkbox"/> POSITIEF (AC-1 t/m AC-28)	Hyden positief voor de nucleair, cytoplasmatisch, mitotisch of niet gefibrillair patroon, gelieve onderstaande labels verder aan te vullen.		

HOMOGEEN (AC-1)

DICHT Fijn gepoeteld (AC-2)

CENTROMEER (AC-3)

GESPOEKELD (AC-4,5)

NUCLEAIR (AC-1 t/m AC-14, AC-29)

NUCLEAIRE DOTS (AC-6,7)

NUCLEOLAIR (AC-8,9,10)

KERNEEMBRAAN (AC-11,12)

PLEOMORF (AC-13,14)

FIBRILLAIR (AC-15,16,17)

FIBRILLAIR (AC-15,16,17)

FIBRILLAIR filamentair (AC-15)

FIBRILLAIR segmentaal (AC-17)

CYTOPLASMATISCH (AC-15 t/m AC-23)

GESPOEKELD (AC-19,20)

RETICULAIR (AC-21)

GOLGI-KICHTIG (AC-22)

STAAPJES en RINGEN (AC-23)

MITOTISCH (AC-24 t/m AC-28)

NIET GEDEFINIËRD (AC-30)

TOEGEGANG (AC-29)

Fijn gepoeteld (AC-4)

Groot gepoeteld (AC-5)

Multipel nucleair site (AC-6)

Enkele nucleair dots (AC-7)

Nucleolair homogeen (AC-8)

Nucleolair gepoeteld (AC-8)

Nucleolair gepoeteld (AC-10)

Kernembran glas (AC-11)

Kernembran gepoeteld (AC-12)

PCNA-achtig (AC-13)

CENPF-achtig (AC-14)

Fibrillair lineair (AC-15)

Fibrillair filamentair (AC-15)

Fibrillair segmentaal (AC-17)

Discrete dots (AC-18)

Dicht fijn gepoeteld (AC-19)

Fijn gepoeteld (AC-20)

Centrosomen (AC-24)

Spindelvezels (AC-25)

Intercellulaire bruggen (AC-27)

Chromosomaal (AC-28)

Nucleair-achtig (AC-30)

CUT-OFF: 1/1 TITER: 1/1 Niet uitgevoerd

Survey: ICAP

ANA IIF pattern survey

Study Objective

Current laboratory and clinical practices in reporting and interpreting ANA patterns in Belgium.

& beyond



Materials and methods

- **International Consensus on Antinuclear Antibody Patterns (ICAP)**



- Consensus ANA pattern definition
- Level of competency (competent/expert) to identify them

- **Questionnaires**

- Content: reporting, familiarity, clinical significance of patterns
- Scope: Belgian laboratories licensed to perform ANA IIF testing Belgian rheumatologists



Van Hoovels et al. *Autoimmun Highlights* (2020) 11:17
<https://doi.org/10.1186/s13317-020-00139-9>

Autoimmunity Highlights

ORIGINAL RESEARCH

Open Access



Current laboratory and clinical practices in reporting and interpreting anti-nuclear antibody indirect immunofluorescence (ANA IIF) patterns: results of an international survey

Lieve Van Hoovels^{1,2*}, Sylvia Broeders³, Edward K. L. Chan⁴, Luis Andrade⁵, Wilson de Melo Cruvinel⁶, Jan Damoiseaux⁷, Markku Viander⁸, Manfred Herold⁹, Wim Coucke³, Ingmar Heijnen¹⁰, Dimitrios Bogdanos¹¹, Jaime Calvo-Alén¹², Catharina Eriksson¹³, Ana Kozmar¹⁴, Liisa Kuhl¹⁵, Carolien Bonroy^{16,17}, Bernard Lauwerys^{18,19}, Sofie Schouwers²⁰, Laurence Lutteri²¹, Martine Vercammen^{22,23}, Miroslav Mayer²⁴, Dina Patel²⁵, William Egner²⁵, Kari Puolakka²⁶, Andrea Tesija-Kuna¹⁴, Yehuda Shoenfeld^{27,28}, Maria José Rego de Sousa²⁹, Marcos Lopez Hoyos³⁰, Antonella Radice³¹ and Xavier Bossuyt^{2,32}

Survey: ANA methodology

A SURVEY ON METHODOLOGICAL ASPECTS OF ANTI-NUCLEAR ANTIBODY TESTING

The use of Indirect Immunofluorescence for
ANA testing in Belgium

Vercammen M, Broeders S, Bonroy C, Coucke W, Lutteri L, Schouwers S,
Van Hoovels L, Bossuyt X. Belgian EASI team - [Sciensano](#)

14th Dresden Symposium on Autoantibodies

.be



DE GRUYTER

Clin Chem Lab Med 2023; 61(7): 1199–1208

EFLM Paper

Martine Vercammen, Carolien Bonroy, Sylvia Broeders, Edward K.L. Chan, Nicola Bizzaro, Dimitrios P. Bogdanos, Luis Andrade, Wim Coucke, Wilson de Melo Cruvinel, Ana Kozmar, Liisa Kuhi, Laurence Lutteri, Maria Jose Rego de Sousa, Sofie Schouwers, Lieve Van Hoovels and Xavier Bossuyt*, on behalf of the EFLM Working Group on Autoimmunity Testing

Analytical aspects of the antinuclear antibody test by HEp-2 indirect immunofluorescence: EFLM report on an international survey

09:55 – 10:20 Detection of antinuclear antibodies: recommendations from EFLM, EASI and ICAP: QA & verification, a practical approach

Martine Vercammen (Department of Laboratory Medicine, AZ Sint-Jan, Brugge, Belgium)

Standardisation in NIS

Autoimmune diagnostics: complex!!!

- Different kits/providers
- Different methodologies -> Difference in results!!
- Different specificities (Ag source, avidity,...) => **Standardisation is needed!!**
-

Standardisation of PR3-ANCA and MPO-ANCA:
evaluation of certified reference materials

Xavier Bossuyt ¹, Doreen Dillaerts,¹ Michael Mahler ²,
Dirk Roggenbuch,^{3,4} Ulrich Leinfelder,⁵ Friederike Hammar,⁵
Wolfgang Schlumberger,⁶ Nina Olschowka,⁷ Jan Damoiseaux ⁸

=> Taken together, using the certified reference material for PR3-ANCA and MPO-ANCA squeezes test results within the same order of magnitude. However, depending on the assays compared, differences and poor correlation between individual results remain.

UKNEQAS collaboration:

- SN/18676: AC-5, dsDNA neg, RNP
 - SN/18677: AC-5
 - SN/18647: Anti-DFS70/LEDGF-p75 (anti-DFS70)
- > Can we standardize?

Harmonisation in NIS

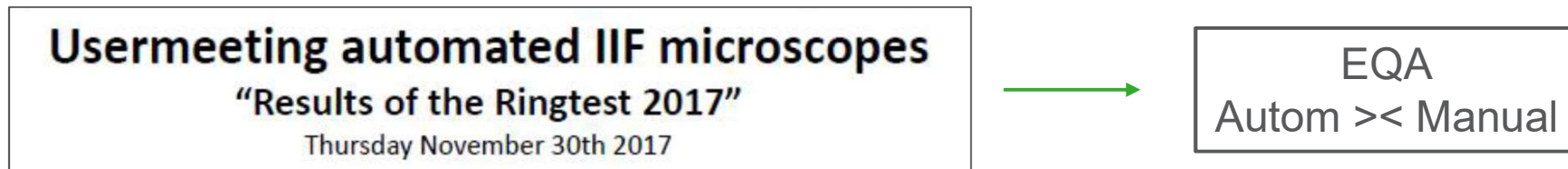
Is this more feasible?

ICAP nomenclature -> IIF aspect reporting

Ring test:

- SN/15085: (fine) speckled, SSA/SSB
- SN/15086: negative

-> Use of automated microscope helps in harmonisation ?



General: test result interpretation < universal way of test reporting

-> Based on cut-off: establishment?

Harmonisation in NIS

Use of Likelihood ratio for test result reporting?!

11:00 - 11:30 **LR in autoimmune diagnostics: basic principles**

Xavier Bossuyt (Department of Microbiology, Immunology and Transplantation, KU Leuven, Leuven, Belgium; Department of Laboratory Medicine, University Hospital Leuven, Leuven, Belgium)

-> Survey BE + EASI 2023



Current application and interest of likelihood ratio reporting in autoimmune serology: results of a Belgian and EASI forum survey



Lieve Van Hoovels^{1,2}, Sylvia Broeders³, Jan Damoiseaux⁴, Wim Coucke³, Carolien Bonroy^{5,6}, Sylvie Goletti⁷, Laurence Lutteri⁸, Sofie Schouwers⁹, Martine Vercammen^{10,11}, Xavier Bossuyt^{2,12}
on behalf of the Belgian expert committee on non-infectious serology of Sciensano and the Belgian EASI working group

¹OLV Hospital Aalst, Department of Laboratory Medicine, Aalst, Belgium; ²Department of Microbiology, Immunology and Transplantation, KU Leuven, Leuven, Belgium; ³Sciensano, Quality of laboratories, Brussels, Belgium; ⁴Central Diagnostic Laboratory, Maastricht University Medical Center, Maastricht, Netherlands; ⁵Department of Laboratory Medicine, Ghent University, Ghent, Belgium; ⁶Department of Diagnostic Sciences, Ghent University, Ghent, Belgium; ⁷Department of Laboratory Medicine, ULB Bruxelles, Brussels, Belgium; ⁸Department of Laboratory Medicine, CHU Liège, Liège, Belgium; ⁹GZA Hospital, Department of Laboratory Medicine, Antwerp, Belgium; ¹⁰AZ St-Jan Hospital, Bruges, Department of Laboratory Medicine, Bruges, Belgium; ¹¹Research group REIM, Vrije Universiteit Brussel (VUB), Brussels, Belgium; ¹²Department of Laboratory Medicine, UZ Leuven, Leuven, Belgium

16th Dresden Symposium on autoimmune antibodies

11:30 - 11:50 **LR: Results of the 2023 survey and practical implementation in the lab**

Sofie Schouwers (Department of Laboratory Medicine, GZA Hospitals/ZAS, Antwerp, Belgium)

Pipeline

- EQA RF/anti-CCP, ANCA, ANA
- EQA Coeliac Disease
- Survey on APS (BE + NL) -> EQA?

11/03/2024
EQA RF/anti-CCP

13:00 – 13:25 APS new classification criteria and daily practice in laboratory diagnosis of APS Katrien Devreese (Coagulation Laboratory, Department of Diagnostic Sciences, Ghent University Hospital, Ghent, Belgium)

13:25 – 13:50 Method comparison in Antiphospholipid antibody measurement Lieve Van Hoovels (Department of Laboratory Medicine, OLV Ziekenhuis, Aalst, Belgium; Department of Microbiology, Immunology and Transplantation, KU Leuven, Leuven, Belgium)

- Publications
 - LR application on EQA results
 - ...
- Alternative 'tool' for the NIS EQA on paper

ACKNOWLEDGEMENTS

Expert committee NIS
RBSLM

QUESTIONS?



Contact

Broeders Sylvia • sylvia.broeders@sciensano.be • +32 2 642 52 25

Presentations, which are allowed, will be published on the Sciensano website